CarbaCamp

Assessment of phenotypic carbapenem susceptibility and genomic epidemiology of *Campylobacter* from animal, food and human domains

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2018	2020	Feb. 2023	Mar. 2023
Evidence of carbapenem non-susceptibility in <i>Campylobacter</i> in humans • Hagiya et al.,2018 • Lehours et al. 2018 Resistance rates for ertag	ETP resistance included in the AMR monitoring of <i>Campylobacter</i> , by the Commission Implementing Decision (EU) 2020/1729 • Range 0.125 – 4 mg/L • ECOFF = 0.5 mg/L Deenem in <i>C. coli</i> (R>0.5)	Observations from the Austrian Agency for Health and Food Safety (AGES) shared with the EURL-AMR: • High rates for ETP non- susceptible Campylobacter from food animals • Possible different wild-type distributions between C. jejuni and C. coli. 30 16 16 10 10 10 10 10 10 10 10 10 10	 EURL-AMR organized a webinar with the EURL network, EFSA and ECDC. Data discussed within the network and: Dr. Philippe Lehours, University of Bordeaux, PU-PH, Bordeaux, France, an expert in AMR as well as with EFSA ECDC
60.0% 50.0% 40.0% 20.0% 20.0% <= 0.12 0.25 0.5 1 2 4 >4 • C. jejuni • C. coli	%R=68.8 %R=68.8 %R=68.8 %R=63.0 %R=63.0 %R=63.0 %R=63.0 %C = 0.12 0.25 0.5 1 2 4 >4 %C = 0.12 0.5 1 2 4 >4 %		
Broiler meat 2022, C. jejuni n=91, C. coli n=54 60,0% 40,0% 30,0% 20,0% 10,0% <= 0.12 0.25 0.5 1 2 4 >4 = C. jejuni $=$ C. coli	$%R=79.6 \qquad \qquad$		
Courtesy of Sunara Roeben-Selo wan <u>sanara-bit guia. Roeben 1-felov can (auges. au</u>			



Main questions emerged from the webinar:

- Is the present ECOFF for ETP set correctly ?
- Is ETP the best carbapenem for the tests ?
- What is the effect of using EUCAST versus CLSI recommended media for AST?
- Are there any differences in the wild-type populations between animal types and between species?
- Can a resistance mechanism be identified as responsible for the higher MIC levels ?



Project structured in 6 "Tasks"







- \Rightarrow strain collections from interested NRLs of the EURL AR network providing data to the EFSA surveillance of AMR.
- \Rightarrow NRLs that have submitted most Campylobacter data of the four animal and food domains to the EFSA surveillance of AMR will be targeted (Survey)
- ⇒ The animal and food strain collection will be based obtaining a representative diversity of strains representing different MIC distributions against ETP ensuring isolates across the MIC range of 0.125 – 4 mg/L.
- $\,\Rightarrow\,$ MTAs between DTU and NRLs

All strains shipped to DTU for analysis

Task 2 – AST by DD



Meropenem

- Ertapenem
- Imipenem
- Pefloxacin (screening for fluoroquinolone resistance)
- Erythromycin (screening for macrolid-resistance)
- Tetracycline.

 \Rightarrow *C. jejuni* strain ATCC 33560 will be used for quality control.

=> EUCAST DD protocol for *Campylobacter* (https://www.eucast.org/ast_of_bacteria/disk_diffusion_methodology) using Muller Hinton agar (MHF (EUCAST recommendation)) of two different suppliers e.g. Oxoid and BioRad

Task 3- AST by broth microdilution



- => AST by broth microdilution
- => Custom sensititre plate design 500 plates minimum order
- \Rightarrow Ertapenem, imipenem, and meropenem
- \Rightarrow range from 0.002 to 32 mg/L.
- => Half a plate for one strain => 300 plates for 600 isolates
- => MH-F (EUCAST recommendation)) of two different suppliers every second week following the EUCAST testing principle

test a subset of the same isolates using the MH+LHB (CLSI recommendations) broth in comparison to MH-F



Task 4&5- WGS and in silico analysis



Illumina technology using the standard EURL-AR WGS protocol

https://www.eurlar.eu/CustomerData/Files/Folders/34 -wgs/628_protocol-for-wgs-v2-2.pdf

- presence of known antimicrobial determinants e.g. *bla*OXA and resistance mutations *por*A or *cme*ABC
- relevant epidemiological markers, and phylogeny using both a gene by gene; cgMLST and SNP approach based on the experience of the EFSA GenCamp project (https://www.efsa.europa.eu/en/supporting/pub/en-1398).
- In addition, the data will be further investigated to detect novelties explaining the non-susceptibility to carbapenems.
- All raw data of the study will be supplemented with additional *C. jejuni* and *C. coli* genomes from ENA



- To determine the wildtype MIC distribution between *C. jejuni* and *C. coli* as well as between the four animal domains and in humans by disk diffusion and broth microdilution.
- To determine the comparability between the EUCAST and CLSI recommended media for MIC determination of *Campylobacter*.
- To determine the genomic diversity of susceptible and nonsusceptible *C. jejuni* and *C. coli* across the different animal species and humans in EU.
- To determine which of ertapenem, imipenem and meropenem would be most suitable for the monitoring of *Campylobacter*.
- To investigate potential resistance mechanism conferring nonsusceptibility to carbapenems as well as the role of blaOXA genes observed in *Campylobacter*.